

Green Synthesis Of Manganese Dioxide And Copper Oxide Nanoparticles Using Phaseolus Lunatus Flower Extract And Evaluation Of Their Antimicrobial Activity

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ABSTRACT

Manganese dioxide nanoparticles (MnO₂-NPs) and copper oxide nanoparticles (CuO-NPs) with suitable surface chemistry possess various beneficial properties that can be used in numerous biomedical applications, including magnetic resonance imaging contrast enhancement, tissue regeneration, hyperthermia, drug delivery, and cell separation. The study discovered that undiscovered Phaseolus Lunatus flower extract can effectively synthesize manganese dioxide nanoparticles (MnO₂-NPs) and copper oxide nanoparticles (CuO-NPs). The nanoparticles were analyzed by UV-visible spectrophotometer, SEM, EDS, XRD, and FT-IR to determine their properties. The synthesized MnO₂-NPs and CuO-NPs were naturally stabilized, cubic in shape, and sized between 26.98 nm and 65.20 nm. The phytochemicals in the leaf act as reducing agents, aiding in the environmentally friendly synthesis of MnO₂-NPs and CuO-NPs with improved antibacterial properties. The functional groups found on the nanoparticles are primarily -OH and -COOH, as indicated by FT-IR analysis, which makes them hydrophilic. Therefore, no additional functional modifications are required for their uses. The synthesized MnO₂ and CuO were tested for their antibacterial activity against Gram-negative (*E. coli* and *P. aeruginosa*) and Gram-positive (*S. aureus* and *B. subtilis*). The zone of inhibition measured 17 mm for *E. coli* and 15 mm for *S. aureus*. Naturally stabilized Gram-negative nanoparticles with herbal properties can be used in a variety of biological applications.

Keywords: MnO₂-NPs, CuO-NPs, Green synthesis, Phaseolus Lunatus, Antimicrobial activity

1. INTRODUCTION

Using plant flower extracts for synthesizing manganese and copper oxide nanoparticles provides eco-friendly and compatible options for pharmaceutical and biomedical applications since they avoid the use of harmful chemicals in the synthesis process [1-2]. Chemical synthesis processes can introduce hazardous compounds that may have negative effects on medical applications. Currently, the biological production of nanoparticles using plant extracts is being utilized by researchers [3,4]. There are numerous physical, chemical, biological, and hybrid ways of synthesizing various types of nanoparticles [5]. Each approach produces nanoparticles with distinct features. Plant-mediated production of metal nanoparticles is currently in progress [6-7]. Green nanotechnology is a field that has garnered significant interest and encompasses many methods aimed at minimizing or eradicating harmful compounds to help improve the environment. The manufacture of metal nanoparticles using inactive plant tissue, plant extracts, exudates, and other living plant components is a contemporary method. Green synthesis of nanoparticles uses environmentally friendly, non-toxic, and safe chemicals [8-12].

Plants and microbes produce inorganic materials that are mostly found in nanoscale sizes. Cellular extracts from various biological entities can produce nanoparticles with varying sizes and chemical contents [13-15]. Using various plant parts, predominantly leaves, for the extraction of metal nanoparticles is an efficient and cost-effective method known as green synthesis. Metal ions undergo bio-reduction throughout the synthesis process. The plant extract components reduce iron ions, while water-soluble heterocyclic components stabilize the nanoparticles. Metal salts can be used as a suitable precursor for reducing plant extracts. Green nanotechnology is a field that has garnered significant interest and involves many methods aimed at minimizing or eradicating harmful compounds to improve the environment.

Phaseolus lunatus is a big, softly hairy, round, climbing, or trailing plant found in India and elsewhere [16]. The entire plant is acknowledged to have medicinal properties in traditional medical systems. The fruit is tasty and possesses diuretic, antipyretic, antibilious, tonic, vulnerary, and antiperiodic properties. It can treat blood disorders, muscle soreness, and dry cough. Their phytochemicals contain hydroxyl, carboxyl, and amino functional groups. These groups can act as efficient metal-reducing agents and capping agents, creating a strong coating on metal nanoparticles in a single process. The manufacture of metal nanoparticles using deactivated plant tissue, plant extracts, exudates, and other components of living plants is a contemporary method [17]. Green synthesis of nanoparticles ensures the use of environmentally friendly, non-toxic, and safe chemicals. MnO_2 -NPs and CuO -NPs nanoparticles have diverse biomedical applications, including serving as magnetic beads for trapping bacteria, creating sensors to detect different biological threat agents, and playing a crucial role in the medical field. We have examined the synthesis method of Mn and Cu nanoparticles using green chemistry. This study involved synthesizing iron nanoparticles by using dried powder from *Phaseolus lunatus* flower. MnO_2 and CuO nanoparticle production is summarized due to its significance in industry and the environment. Scientific research has confirmed the pharmacological effects of *Phaseolus lunatus* mentioned in Ayurvedic texts [18-19]. Studies have shown that the plant exhibits strong antioxidant, diuretic, antihyperglycemic, anticancer, analgesic, and antidepressant properties. This work examines the characterization and production methods of iron nanoparticles and evaluates their antibacterial efficacy. The MnO_2 -NPs and CuO -NPs nanoparticles were synthesized by using potassium permanganate and copper chloride as the manganese and copper sources and *Phaseolus lunatus* flower extract as the reducing agent and stabilizer.

2. EXPERIMENTAL

2.1. Collection of Plant Material and Preparation of Extract

Flowers of *Phaseolus lunatus* were gathered from Local farmers in, the district of Maharashtra, India. The fresh flowers were used for all testing methods.

2.2. Chemicals

The investigation used potassium permanganate (KMnO_4) and copper chloride hexahydrate ($\text{CuCl}_2 \cdot 6\text{H}_2\text{O}$, 98%) and a solvent of the highest purity and analytical grade obtained from S. D. fine chem.

2.3. Extraction Preparation

The *Phaseolus lunatus* flower components were cleaned and kept at a temperature of -4°C . To produce the extract, about 5 g of ground, air-dried *Phaseolus lunatus* samples (flower) were boiled with 100 ml of double-distilled water in an Erlenmeyer flask while being agitated constantly for 15 minutes. The extract was cooled to ambient temperature, filtered, and then stored at -4°C for future use.

2.4. Preparation of MnO_2 -NPs

Manganese dioxide nanoparticles were prepared using an adjusted procedure based on earlier research studies [20-25]. Simply, mix a 0.01 M solution of KMnO_4 with the *Phaseolus lunatus* flower extract in a 1:1 volume ratio. MnO_2 -NP nanoparticles were promptly prepared using the reduction procedure. The mixture was agitated for 60 minutes and then left at room temperature for an additional 30 minutes to achieve a colloidal suspension. The mixture was centrifuged, washed with ethanol multiple times, and then dried at 40°C under a vacuum to produce the MnO_2 -NPs. Flowers of *Phaseolus lunatus* have superior reduction capability against KMnO_4 , as indicated by the exterior color change. Flowers meeting the criteria were chosen for additional processes. Following the confirmation test, the MnO_2 -NPs were synthesized using the same process for additional characterization.

Preparation of CuO -NPs

A modified protocol based on previous research work was used to manufacture copper oxide nanoparticles [26-28]. To begin, just combine the *Phaseolus lunatus* extract in a 1:1 volume ratio with a 0.01 M solution of $\text{CuCl}_2 \cdot 6\text{H}_2\text{O}$. Rapid production of CuO nanoparticles was achieved by the reduction process. To obtain a colloidal suspension, the liquid was stirred for 60 minutes and then allowed to stand at room temperature for a further 30 minutes. To prepare the CuO nanoparticles, the mixture was centrifuged, repeatedly cleaned with ethanol, and vacuum-dried at 40°C . The external color shift indicates that the *Phaseolus lunatus* flower have a higher reduction capability against copper chloride than other plant components (seeds and fruit). Flowers

that satisfied the requirements were selected for the next steps. The CuO-NPs were generated using the same procedure for further characterization after the confirmation test.

2.5. Characterization techniques

Characterization techniques aid in accurately and rapidly understanding the unique properties of substances or nanocrystals being researched, providing reliable insights into the measured values. The synthesized MnO_2 and CuO nanoparticles underwent various characterization studies to analyze specific properties including structural, morphological, elemental composition, particle size, and functional groups. These properties were precisely examined using advanced techniques such as UV-VIS spectroscopy (JASCO V650 UV-Vis model), XRD (PANalytical X'Pert Pro instrument with Cu $\text{K}\alpha_1$ radiation of wavelength (λ) of 1.5406 ($^\circ\text{A}$)), SEM, (FEI Nova Nano SEM 450) and EDS (Bruker XFlash 6I30) and FT-IR (BRUCKER). The strategies were useful in confirming that our method is well-optimized and meets the requirements.

2.6. Antimicrobial Activity Test

The antimicrobial ability of synthesized MnO_2 and CuO nanoparticles was examined to investigate the herbal functionality of nanoparticles.

Antimicrobial activity was assessed using the disc diffusion method, as described in a previous study [29-30]. The organisms employed for the antibacterial test were: Gram-negative: *E. coli*; and Gram-positive. *Staphylococcus aureus* Standard drug reference - Streptomycin at a concentration of 20 $\mu\text{g/ml}$.

3. RESULTS AND DISCUSSION

A single-step process coats metal nanoparticles and causes a color change from yellowish brown to brownish black. The color shift confirmed the formation of MnO_2 -NPs and CuO-NPs. The flowers of the Phaseolus lunatus plant exhibit a superior ability to generate MnO_2 and CuO nanoparticles compared to other components of the plant including seeds and fruits.

3.1. UV-visible spectra

The preface test compares the UV-visible absorption peak at 516 and 713 nm, showing plant detritus and protein-bound MnO_2 -NPs and CuO-NPs respectively [31] (**Figure 1**). The discrepancy observed may be due to chloride interference in the plant extract [32].

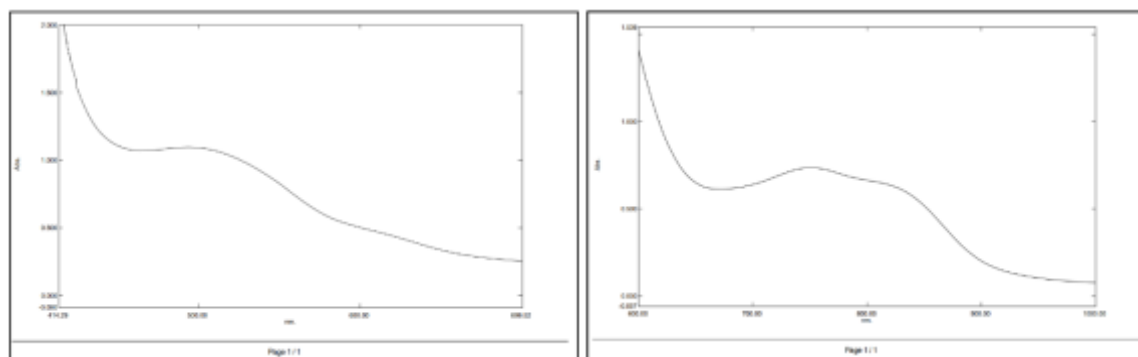


Figure 1. UV-vis spectra of MnO_2 -NPs and CuO-NPs

3.2. EDX spectra

The EDX spectrum in **Figure 2** indicates the presence of chemicals such as Chloride, Calcium, and Potassium in the reaction mixture together with Mn and Cu nanoparticles, suggesting the formation of a complex with the plant extract.

The analysis showed that the flower extract from Phaseolus Lunatus contains 30.18% manganese and 34.19% oxygen in the total weight of MnO_2 -NPs and 19.77% copper and 34.46% oxygen in the total weight of CuO-NPs. This is derived from the bremsstrahlung X-ray intensity concerning energy. Carbon alters the enzyme molecule's physical structure, revealing the suitable chemically active spots for reaction. Carbon neutralizes organic anions and other chemicals in the plant, stabilizing pH between 7 and 8, which is ideal for enzyme activity.

The concentration of carbon (C) in the cell dictates the number of enzymes that can be activated and the speed at which chemical processes can occur.

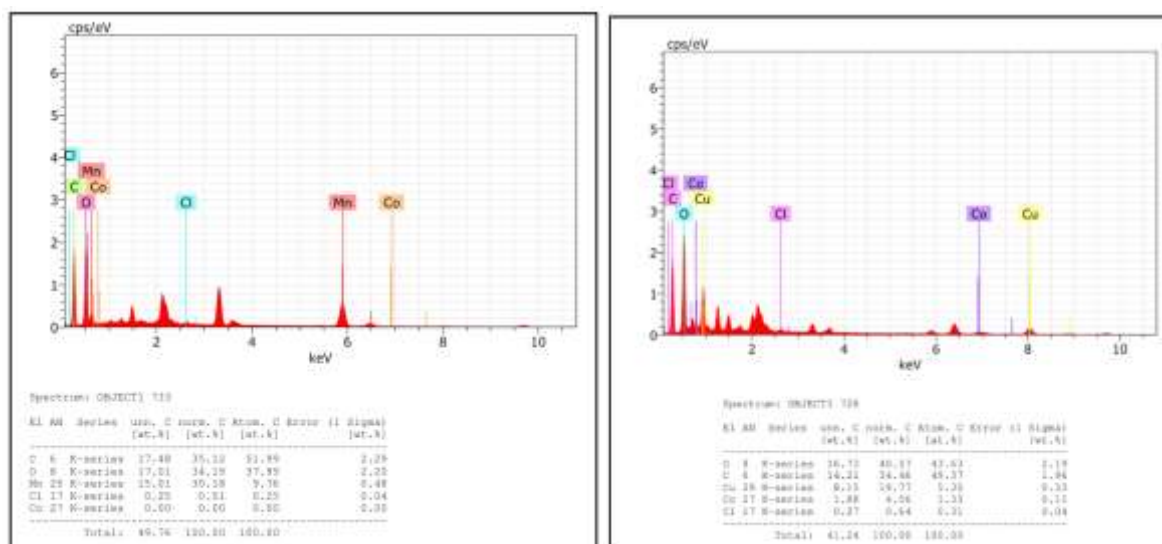


Figure 2: EDAX spectrum for MnO₂-NPs and CuO-NPs

3.3. SEM analysis:

The sample underwent SEM analysis for morphological examination and size measurement. The SEM was used to study the formation of MnO₂-NPs and CuO-NPs and their morphological dimensions. The study showed that the average size of the nanoparticles ranged from 26.98 nm to 65.20 nm, which aligns with findings from earlier studies. The production of cube-shaped manganese and copper nanoparticles is also demonstrated in **Figures 3** and **4**. The preparation of cube-shaped nanoparticles is caused by plant enzymes and chloride. The presence of carbon compounds in the sample influences the morphology of the nanoparticles. SEM micrographs provide a distinctive three-dimensional look that is beneficial for analyzing the surface structure of a material due to the narrow electron beam and great depth of field.

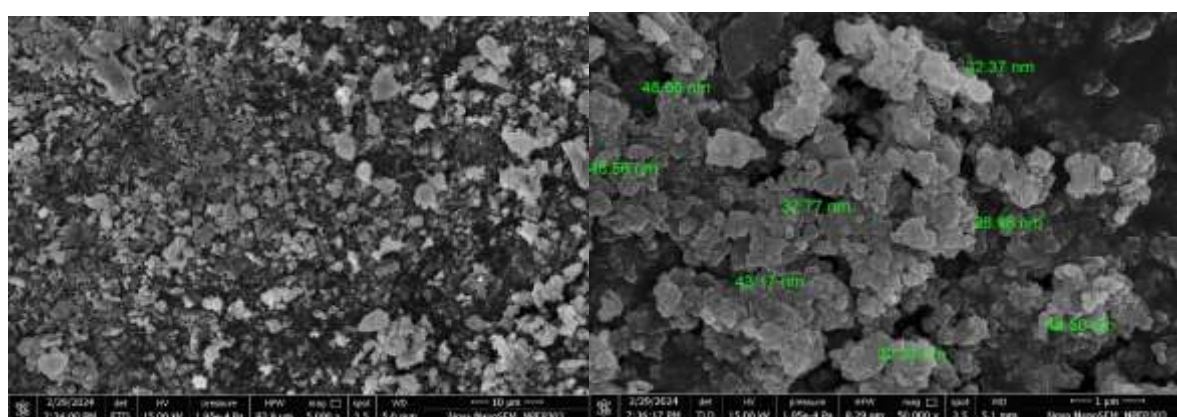


Figure 3: FE-SEM images of MnO₂-NPs with a magnification (10µm and 1µm)

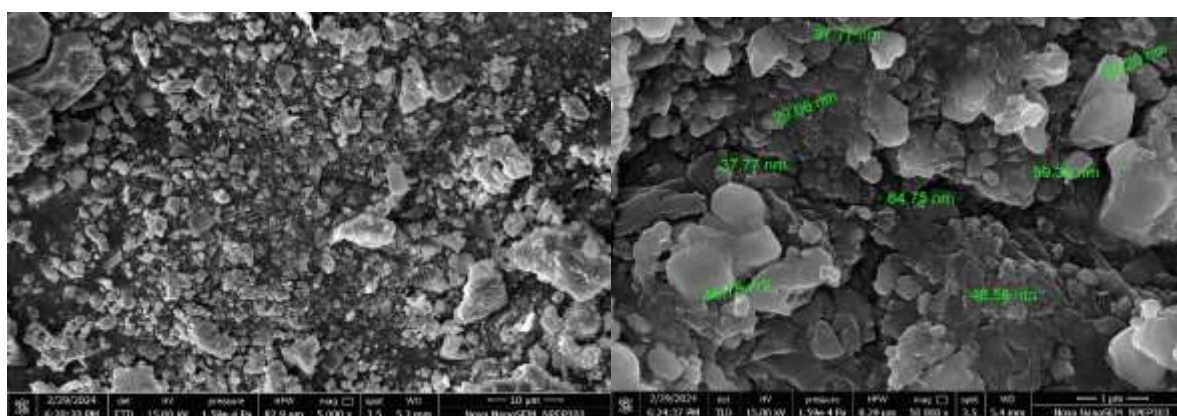


Figure 4: FE-SEM images of CuO-NPs with a magnification (10µm and 1µm)

3.4. X-ray diffraction spectra

Figure 5 displays the X-ray diffraction patterns of the MnO₂-NPs and CuO-NPs synthesized with Phaseolus lunatus extract. The X-ray diffraction analysis revealed strong diffraction peaks at 2θ values of 28.40° and 35.62°, corresponding to the hkl values of 220 and 222, indicating the crystalline phase of MnO₂-NPs and CuO-NPs nanoparticles matching JCPDS card No. 39-1346 and JCPDS card No. 89-4319. The grain size was determined using the Debye-Scherrer equation, establishing a relationship between peak broadening in XRD and particle size. The Scherrer equation was used to determine that the average crystallite sizes of the MnO₂-NPs and CuO-NPs fall within the range of 16 nm to 19 nm. The results showed that all the nanoparticles had a spinel structure with a face-centered cubic phase.

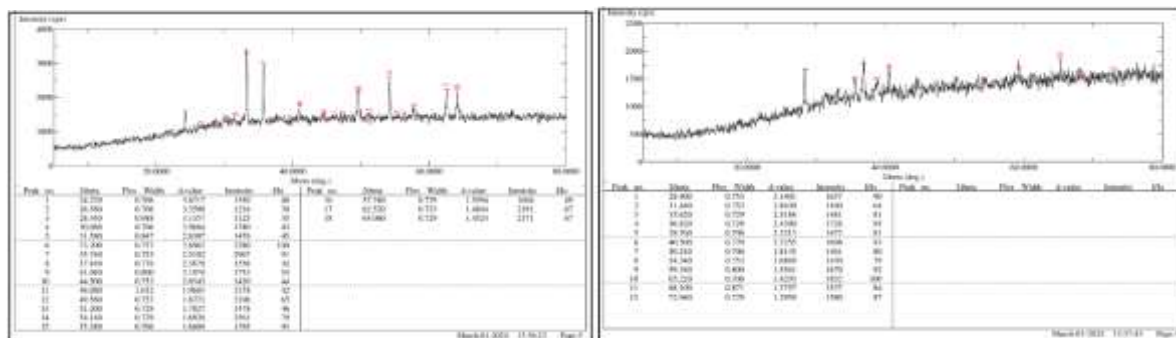


Figure 5: XRD spectra of MnO₂ and CuO NPs

3.5. FT-IR analysis:

FT-IR analysis identified stretching vibrations at 3329-3639 cm⁻¹, 1584-1587 cm⁻¹, and 613-617 cm⁻¹ within the 500-4000 cm⁻¹ area (**Figure 6**). The peaks indicate that the bonding in the sample validates the participation of the reducing agent in the creation of MnO₂-NPs and CuO-NPs. The peak at 3329-3639 cm⁻¹ represents the -OH bond stretching in the aqueous phase and the reduction of metal salts. The peak at 1584-1587 cm⁻¹ corresponds to the C=O bond stretching, indicating the presence of phytochemicals and amino acids that stabilize and act as capping agents in the plant extract. The remaining unexplained peaks indicate a minor number of organic acids that contribute to the low pH of the sample, facilitating the formation of the MnO₂-NPs and CuO-NPs. The prominent peak at 613-617 cm⁻¹ corresponds to the inorganic stretching of MnO₂-NPs and CuO-NPs. The zeta potential of MnO₂-NPs and CuO-NPs was measured at -50 meV, indicating exceptional stability attributed to the significant negative surface charge.

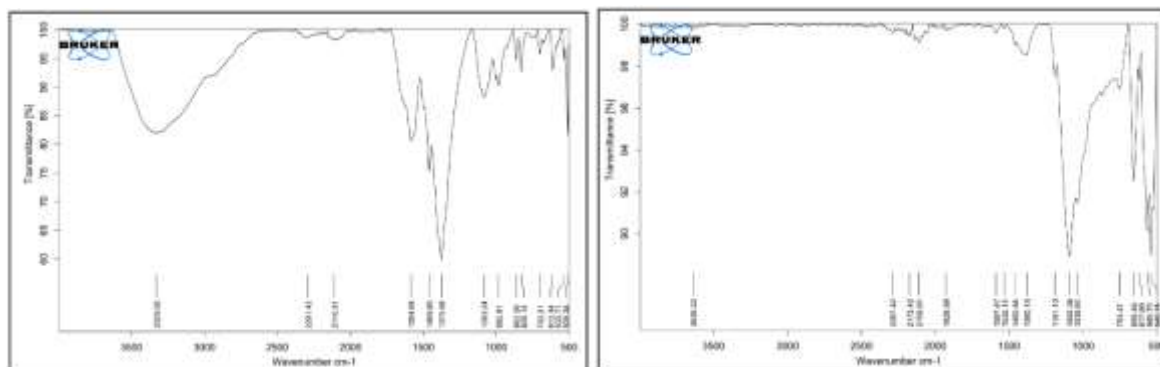


Figure 6: FT-IR spectra of MnO₂ and CuO NPs

The synthesized MnO₂-NPs and CuO-NPs exhibit moderate antimicrobial activity compared to the reference drug. They show higher antibacterial activity against *E. coli* and moderate activity against *S. aureus*. The inhibitory activities of the MnO₂-NPs and CuO-NPs in culture media are comparable to the standard antimicrobial Ampicillin.

Synthesized nanoparticles exhibit equivalent antibacterial efficacy compared to the original plant extract. The plant's herbal property-based functional groups covering the surface of NPs activate antibacterial capabilities. Reduced NP size enhances NP penetration into the cell wall, resulting in cell death. After conducting moderate activity, it was determined that *E. coli* was the more vulnerable bacteria and *S. aureus* was the more resistant. The MnO₂-NPs and CuO-NPs are naturally stabilized and possess a larger surface area suitable for numerous applications. The components involved in the production of MnO₂-NPs and CuO-NPs need to be isolated and purified to prevent the presence of plant contaminants that can affect the properties of the NPs.

3.6. Screening of antibacterial activity of manganese dioxide and copper oxide nanoparticles synthesized by *Phaseolus lunatus* flower extract

The antibacterial properties of MnO₂-NPs and CuO-NPs nanoparticles synthesized through green methods were evaluated against Gram-positive and Gram-negative bacterial strains using the agar well diffusion method. The results revealed that MnO₂-NPs nanoparticles significantly inhibited the growth of *B. subtilis* (22 mm) and *E. coli* (19 mm) at a concentration of 22 µg. In contrast, copper oxide nanoparticles exhibited moderate inhibition of *E. coli* (14 mm) and *S. aureus* (15 mm) at the same concentration. The inhibition zones produced by the commercial antibiotic ciprofloxacin (19 µg/disc) ranged from 10 to 19 mm, surpassing those generated by the nanoparticles. The concentration of synthesized MnO₂-NPs and CuO-NPs nanoparticles played a significant role in determining their antibacterial efficacy. These nanoparticles exhibited notable antibacterial activity against human pathogens such as *E. coli*, *K. pneumonia*, *S. aureus*, and *B. subtilis*.

4. CONCLUSION

The phytochemicals in the sample functioned as capping and reducing agents, naturally stabilizing the NPs. Despite having a lower nanoparticle yield compared to other chemical and physical processes, this method is preferred due to its non-toxic green synthesis and natural functionalization with herbal characteristics. The plant's herbal characteristics are demonstrated by its moderate antibacterial effectiveness against the selected pathogen. Only a small number of NPs can use the plant extract as a reducing agent for synthesizing NPs effectively. Future research will focus on validating the superparamagnetic for its application in cancer therapy.

5. REFERENCES

1. Azhir, E., Reihane, E., Mansour, M., and Parvane, P. (2015) Preparation, characterization and antibacterial activity of manganese oxide nanoparticles. *Physical Chemistry Research*, 3(3), 197-204.
2. Ren, G., Dawei, H., Eileen, C., Miguel, A., Vargas-Reus, P. R., and Allaker, R. P. (2009) Characterization of copper oxide nanoparticles for antimicrobial applications. *International Journal of Antimicrobial Agents*, 33(6), 587-590.
3. Mahdavi, M. (2013) Green biosynthesis and characterization of magnetic iron oxide (Fe₃O₄) nanoparticles using seaweed (*Sargassum muticum*) aqueous extract. *Molecules*, 18(5), 5954-5964. doi: 10.3390/molecules18055954
4. Singh, C. (2011) A Green biogenic approach for the synthesis of gold and silver nanoparticles using zingiber officinale. *Digest J. Nanomat. Biostr.*, 6(2).
5. Akhtar, M. S., Panwar, J., and Yeoung-Sang Y. (2013) Biogenic synthesis of metallic nanoparticles by plant extracts. *ACS Sustainable Chemistry & Engineering*, 1(6), 591-602.
6. Mittal, A. K., Chisti, Y., and Banerjee, U. C. (2013). Synthesis of metallic nanoparticles using plant extracts. *Biotechnology advances*, 31(2), 346-356.
7. Bao, Y., Jian, H., Ke, S., Jie, G., Xianwu, Z., and Shima, L. (2021) Plant-extract-mediated synthesis of metal nanoparticles. *Journal of Chemistry*, 2021, 1-14.
8. Mohamad, N. A. N., Nur Afiah Arham, J. J., and Abdul, H. (2014) Plant extract as reducing agent in the synthesis of metallic nanoparticles: a review. *Advanced Materials Research*, 832, 350-355.
9. Irvani, Siavash. "Green synthesis of metal nanoparticles using plants." *Green Chemistry* 13, no. 10 (2011): 2638-2650.
10. Parsons, J. G., J. R. Peralta-Videa, and J. L. Gardea-Torresdey. "Use of plants in biotechnology: synthesis of metal nanoparticles by inactivated plant tissues, plant extracts, and living plants." *Developments in environmental science* 5 (2007): 463-485.
11. Marchiol, Luca. "Synthesis of metal nanoparticles in living plants." *Italian Journal of Agronomy* 7, no. 3 (2012): e37-e37.
12. Ishak, NAI Md., Kamarudin, S. K., and Timmiati, S. N. (2019) Green synthesis of metal and metal oxide nanoparticles via plant extracts: an overview. *Materials Research Express*, 6(11), 112004.
13. Fierascu, R., Iona, R., and Dumitriu, I. (2010) Noble metals nanoparticles synthesis in plant extracts. *Synthesis*, 1, 22.
14. Yadi, M., Ebrahim, M., Bahram, S., Soodabeh, D., Immi, A., Rovshan, K., Mohammad, N. (2018) Current developments in green synthesis of metallic nanoparticles using plant extracts: a review. *Artificial Cells, Nanomedicine, and Biotechnology*, 46(3), 336-343.
15. Küünal, S., Protima, R., and Erwan, R. (2018) Plant extract mediated synthesis of nanoparticles. In *Emerging applications of nanoparticles and architecture nanostructures*, 411-446.
16. Jean-Pierre, B., Rocha, O., Degreef, J., Maquet, A., and Guarino, L. (2006) *Phaseolus lunatus* L. *Plant resources of tropical Africa (Prota)*, 1.
17. Heil, M. (2004) Induction of two indirect defenses benefits Lima bean (*Phaseolus lunatus*, Fabaceae) in nature. *Journal of Ecology*, 92(3), 527-536.
18. Granito, M., Yannellis, B., and Alexia, T. (2007), Chemical composition, antioxidant capacity and functionality of raw and processed *Phaseolus lunatus*. *Journal of the Science of Food and Agriculture*, 87(15), 2801-2809.

19. Novelo-Cen, L., and Betancur-Ancona, D. (2005) Chemical and functional properties of Phaseolus lunatus and Manihot esculenta starch blends. *Starch-Stärke*, 57(9), 431-441.
20. Moon, S. A., Salunke, B., Bassam, A., Ezhaveni, S., and Beom, S. K. (2015) Biological synthesis of manganese dioxide nanoparticles by Kalopanax pictus plant extract. *IET nanobiotechnology*, 9(4), 220-225.
21. Ogunyemi, S. O., Feng, Z., Yasmine, A., Muchen, Z., Yanli, W., Guochang, S., Wen, Q., and Bin, L. (2019) Biosynthesis and characterization of magnesium oxide and manganese dioxide nanoparticles using Matricaria chamomilla L. extract and its inhibitory effect on Acidovorax oryzae strain RS-2." *Artificial cells, nanomedicine, and biotechnology*, 47(1), 2230-2239.
22. Joshi, N. C., Siddiqui, F., Mohd, S., and Singh, A. (2020) Antibacterial activity, characterizations, and biological synthesis of manganese oxide nanoparticles using the extract of aloe vera. *Asian Pac J Health Sci*, 7, 27-29.
23. Mahlangeni, N. T., Judie, M., Roshila, M., Himansu, B., and Hafizah, C. (2020) Biogenic synthesis, antioxidant and antimicrobial activity of silver and manganese dioxide nanoparticles using Cussonia zuluensis Strey. *Chemical Papers*, 74, 4253-4265.
24. Souri, M., Vahid, H., Nasser, G., and Alireza, S. (2019) Procedure optimization for green synthesis of manganese dioxide nanoparticles by Yucca gloriosa leaf extract. *International Nano Letters*, 9, 73-81.
25. Aarthi, R., and Periyasamy, A. (2023) Biological synthesis of manganese oxide nanoparticles from aerial parts of Prunus dulcis and their in vitro investigation of medical properties." *Applied Organometallic Chemistry*, 37(12), e7283.
26. Rehana, D., Mahendiran, D., Senthil Kumar, R., and Kalilur Rahman, A. (2017) Evaluation of antioxidant and anticancer activity of copper oxide nanoparticles synthesized using medicinally important plant extracts." *Biomedicine & Pharmacotherapy*, 89, 1067-1077.
27. Thakar, M., Jha, S., Khongdet, P., Manne, R., Qureshi, Y., and Hari Babu, V. (2022) X-ray diffraction (XRD) analysis and evaluation of the antioxidant activity of copper oxide nanoparticles synthesized from leaf extract of Cissus Virginia. *Materials Today: Proceedings*, 51, 319-324.
28. Muthuvel, A., Jothibas, M., and Manoharan, C. (2020) Synthesis of copper oxide nanoparticles by chemical and biogenic methods: photocatalytic degradation and in vitro antioxidant activity. *Nanotechnology for Environmental Engineering*, 5, 1-19.
29. Awwad, A. M., Borhan, A. A., and Nida, M. S. (2015) Antibacterial activity of synthesized copper oxide nanoparticles using Malva sylvestris leaf extract. *SMU Med J*, 2(1), 91-101.
30. Asamoah, R. B., Yaya, A., Mensah, B., Nbalayim, P., Apalangya, V., Bensah, Y. D., Damoah, L. N. W., Agyei-Tuffour, B., Dodoo-Arhin, D., and Annan, E. (2020) Synthesis and characterization of zinc and copper oxide nanoparticles and their antibacterial activity. *Results in Materials*, 7, 100099.
31. Bezza, F. A., Shepherd M., Tichapondwa, and Evans, M. (2020) Fabrication of monodispersed copper oxide nanoparticles with potential application as antimicrobial agents. *Scientific Reports*, 10(1), 16680.
32. Singh, J., Gurjas, K., and Rawat, M. (2016) A brief review on synthesis and characterization of copper oxide nanoparticles and its applications. *J. Bioelectron. Nanotechnol*, 1(9).
33. Keabadile, O. P., Adeyemi, O. Aremu, S. E. E., and Omolola, E. F. (2020) Green and traditional synthesis of copper oxide nanoparticles-Comparative study. *Nanomaterials*, 10(12), 2502.